

Sampling and sample storage

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New Zealand Government

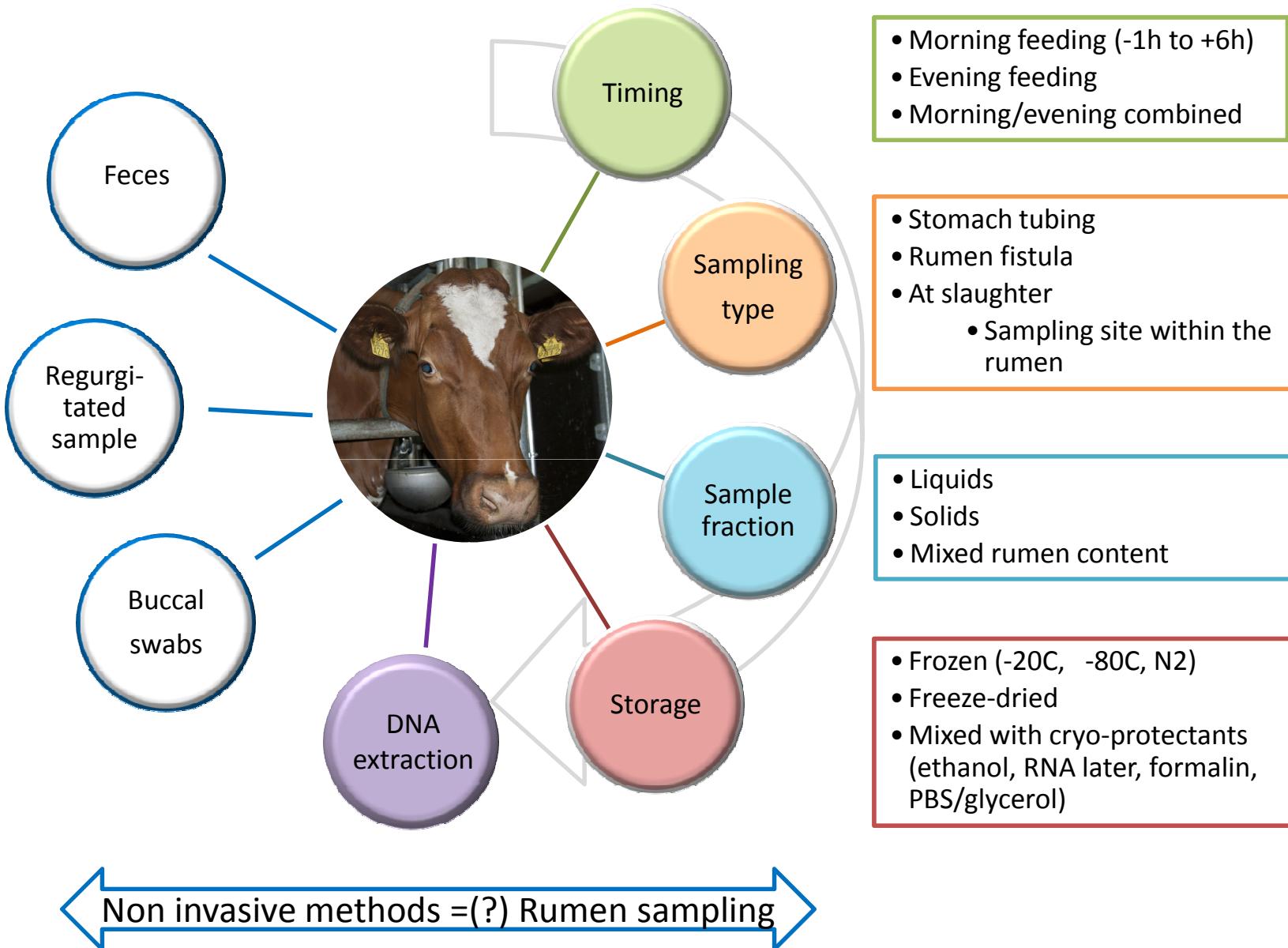


Joint Ruminomics and RMG Network Workshop

22/06/2013, Dublin, Ireland

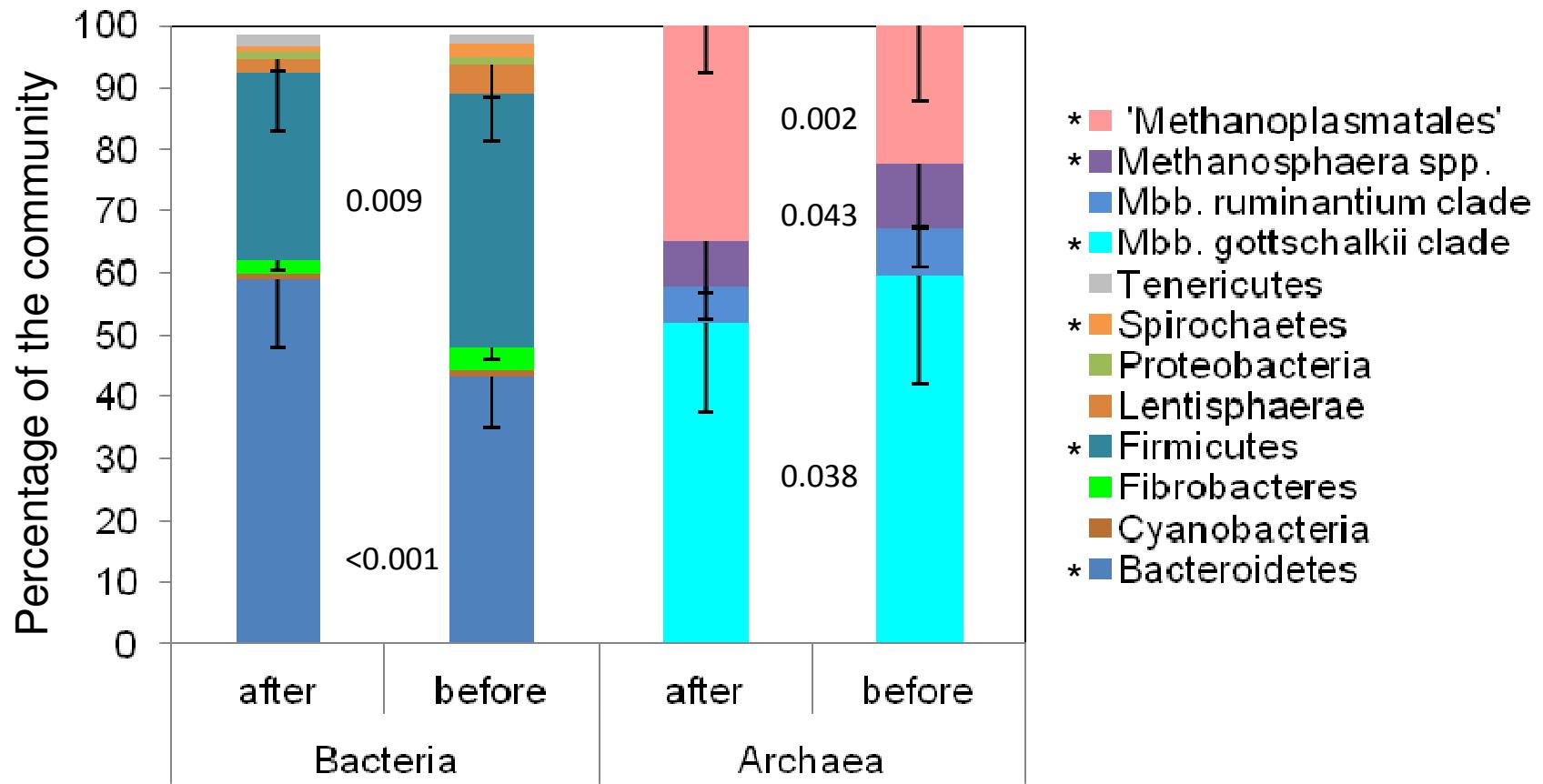


Sampling and storage options



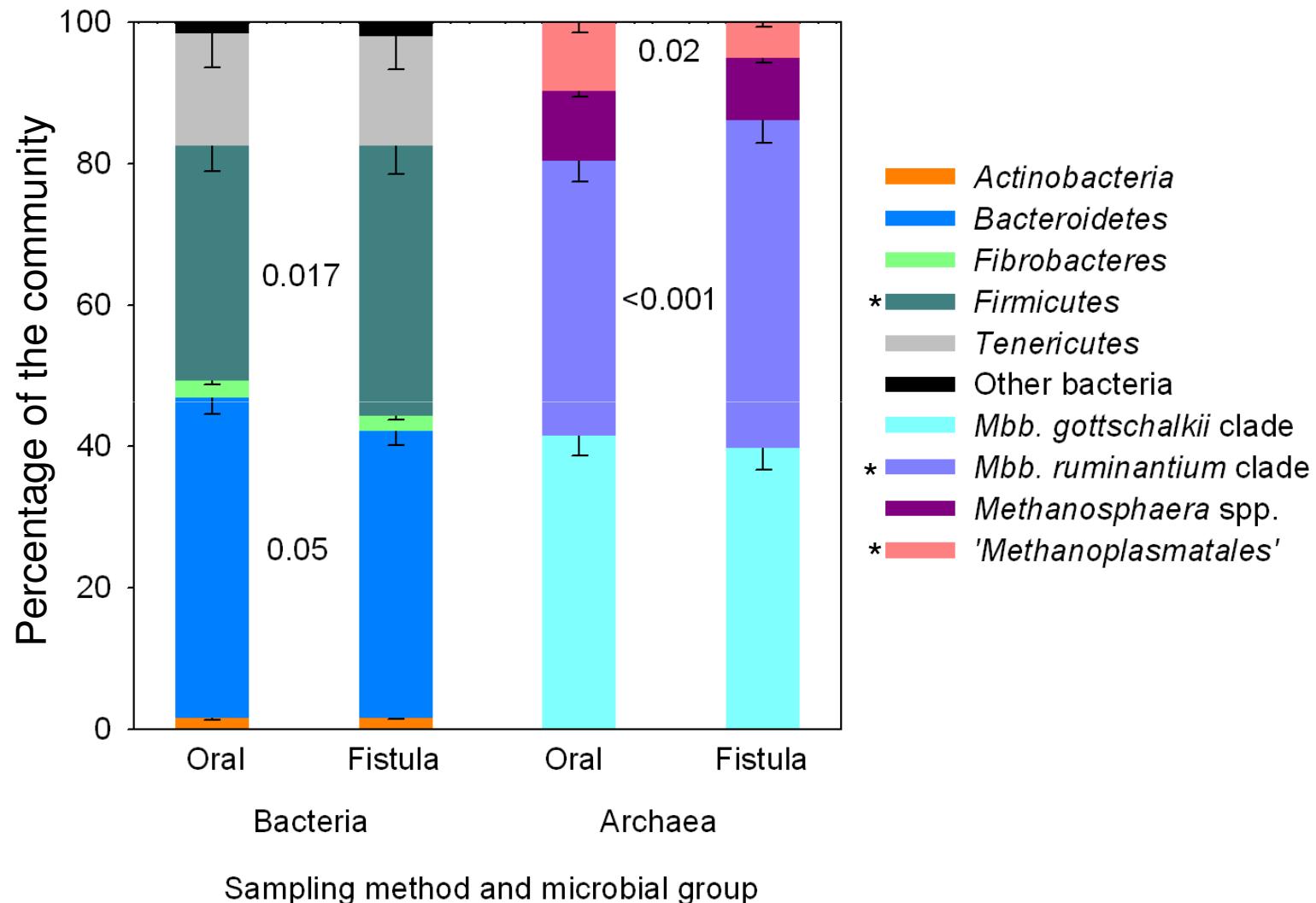
Timing

Before v after feeding



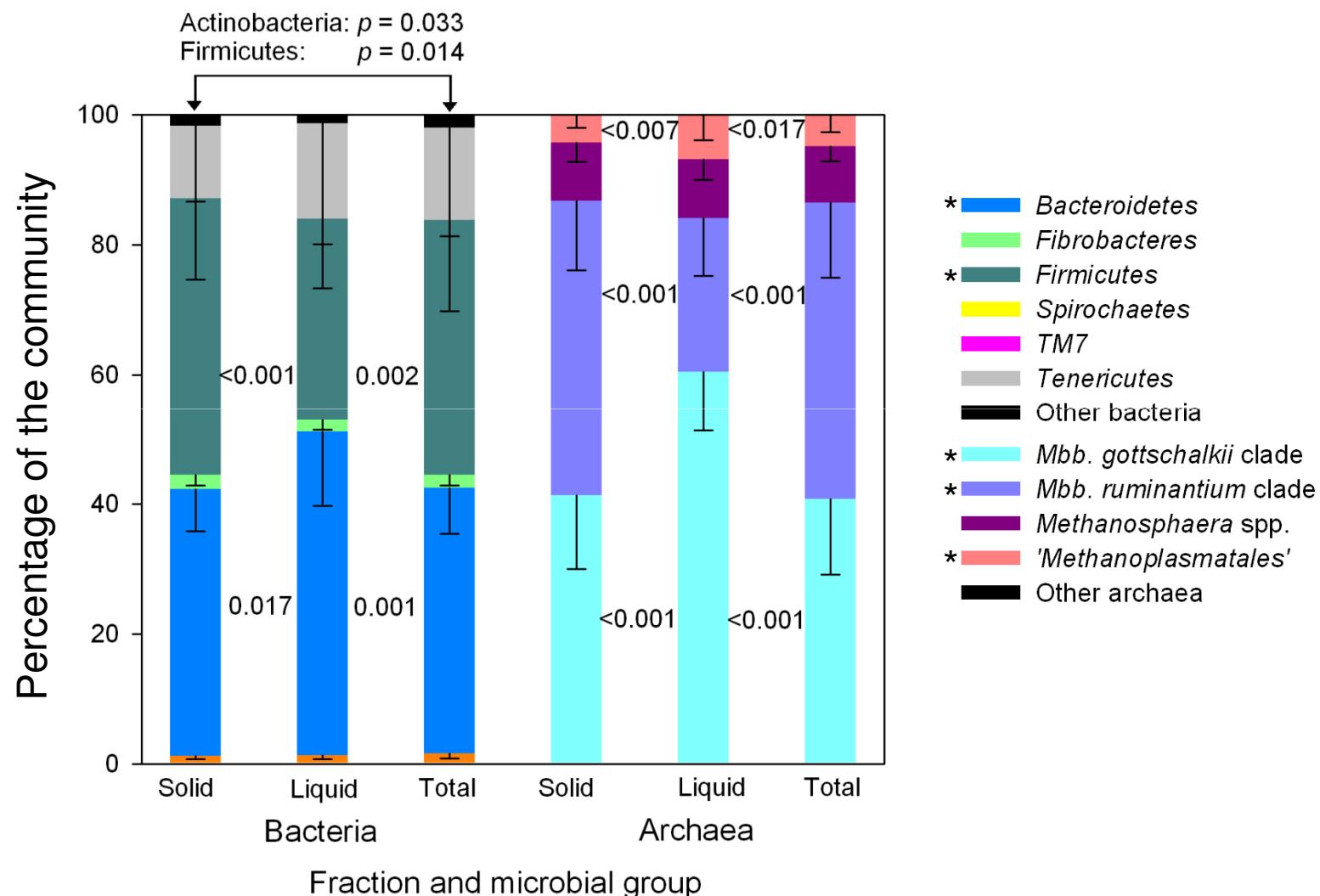


Fistula v stomach tube



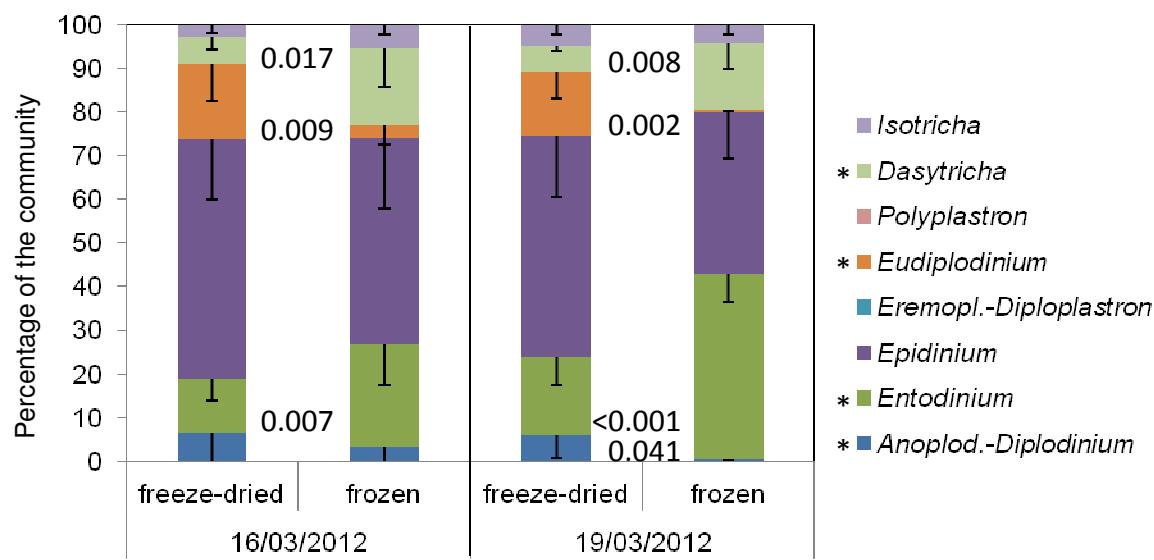
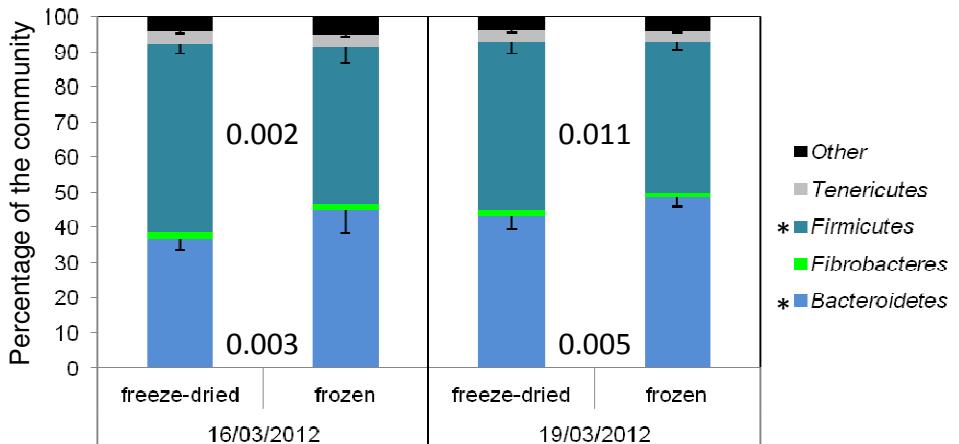
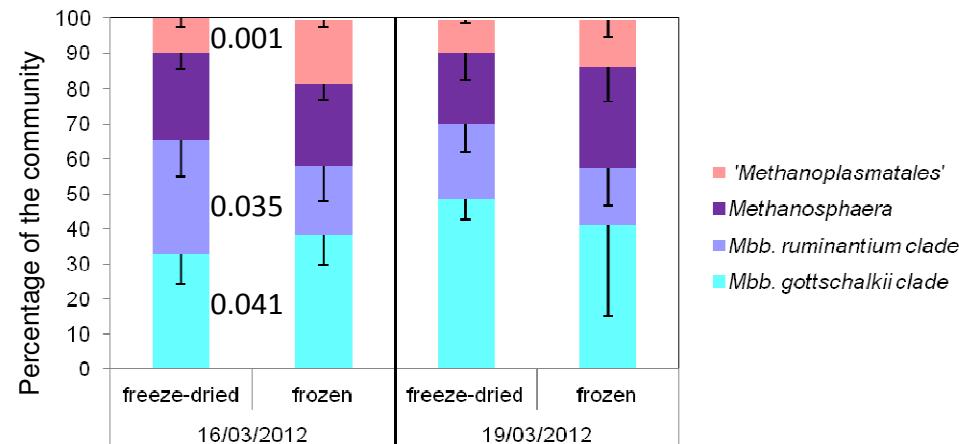


Whole v liquid v solid



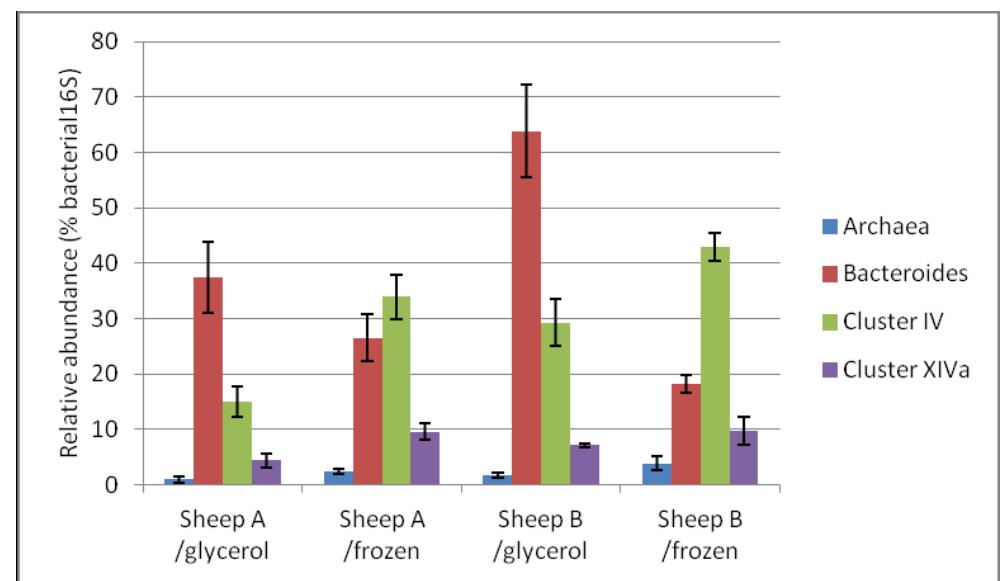
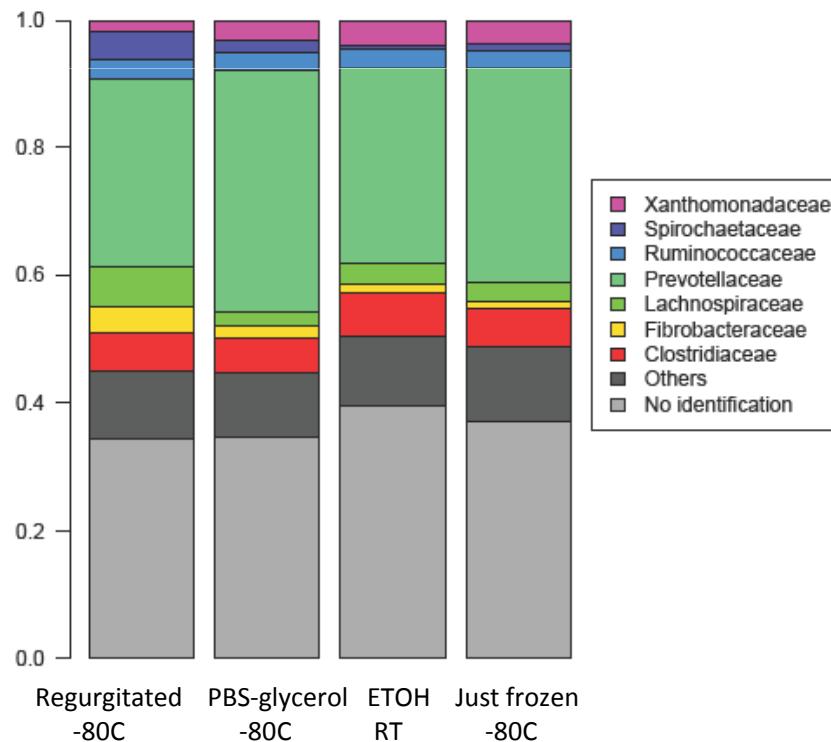
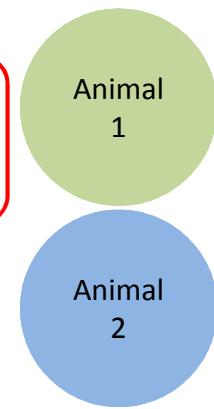


Freeze-dried v frozen only



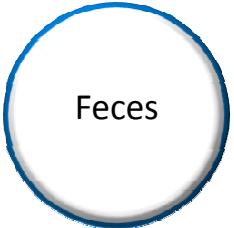


Cryo-protectants

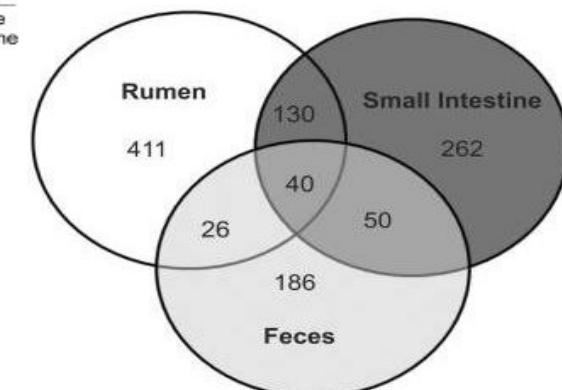
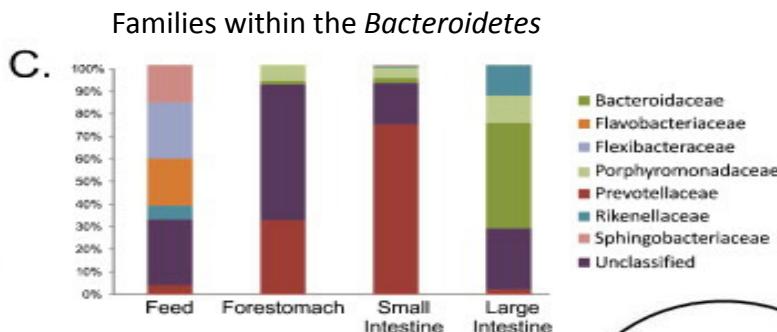
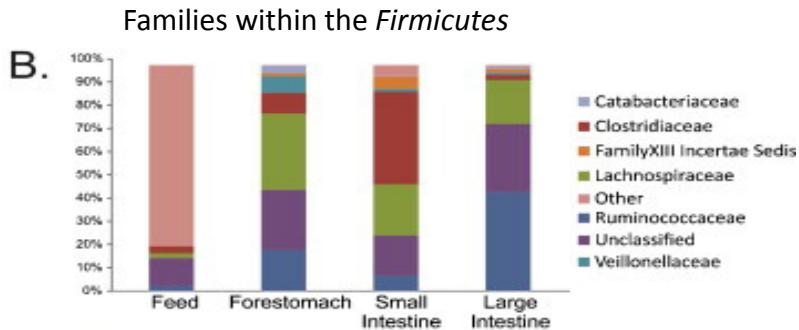
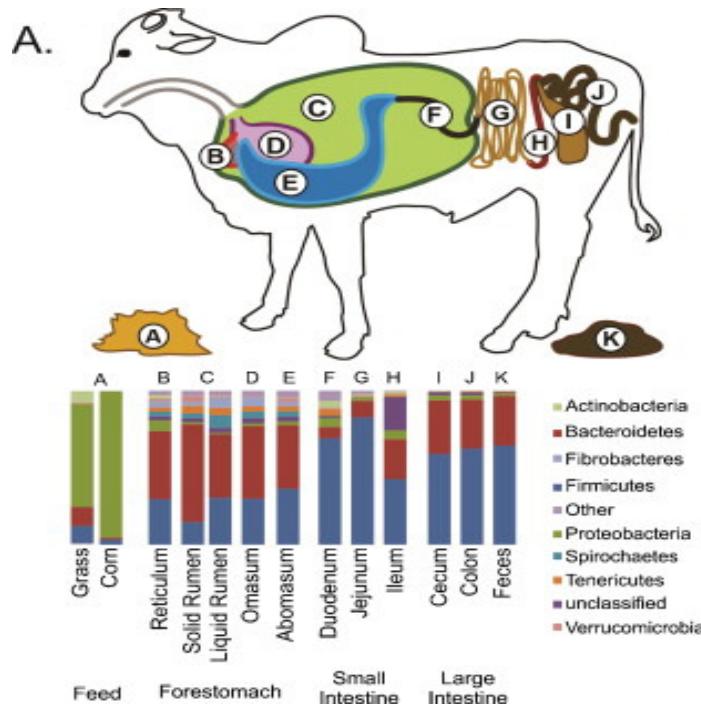


McKain et al. short communication

sheep, n=2



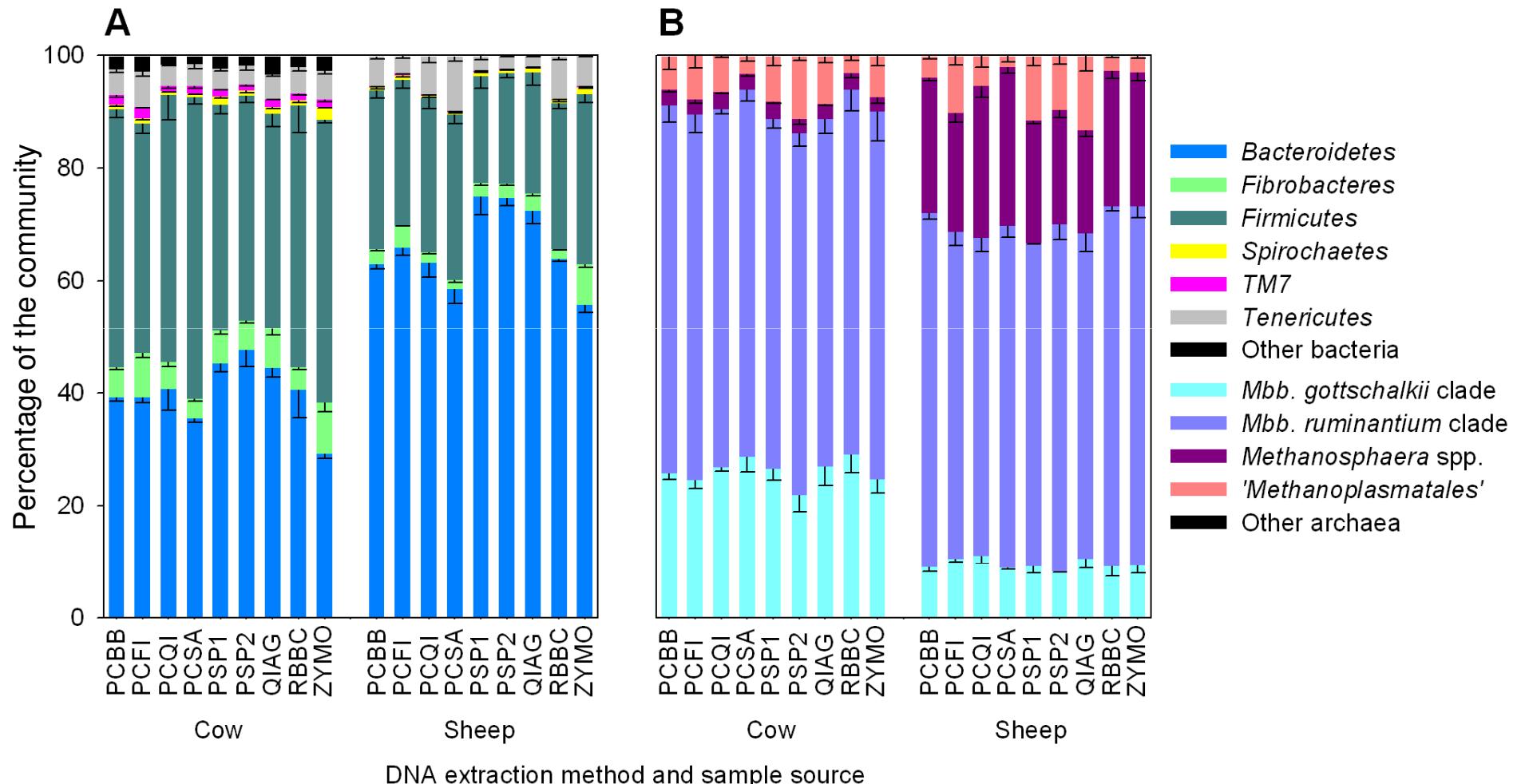
Non-invasive sample collection



de Oliveira et al. 2013 Characterizing the microbiota across the gastrointestinal tract of a Brazilian Nelore steer. Veterinary Microbiology 164: 307-314.



DNA extraction



Take home message

Every sampling technique will represent the rumen microbial community

It is not yet possible to tell which techniques truly represent the rumen microbial community

- There is no “right” or “wrong”

Design your study carefully and be aware of differences introduced by selection of even subtly different protocols

- ...especially if looking for subtle changes

Large cohort studies should strive to use the same or comparable protocols

- Global Rumen Census
- RuminOmics
- ring test

Many more downstream variables to be considered

- Primers, data analysis, taxonomic frameworks, etc.

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Other valuable references:

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